TaKaRa Ex Taq™

Code No. HRR001A Size: 250 units

Shipping at − 20°C Store at − 20°C

Supplied Reagents: 10X Ex Taq Buffer 1 ml dNTP Mixture 800 μ I

Lot No.

Conc.: $units/\mu I$ Volume: μI

Expiration Date:

Storage Buffer: 20 mM Tris-HCl (pH8.0)

100 mM KCI 0.1 mM EDTA 1 mM DTT 0.5% Tween 20 0.5% NP-40 50% Glycerol

Supplied 10X Ex Taq Buffer: Mg²⁺ concentration (10X): 20 mM

Supplied dNTP Mixture:

dNTP Mixture is ready for use in PCR without dilution.

Concentration: 2.5 mM of each dNTP

Form : Dissolved in water (sodium salts), pH 7 - 9

Purity : \geq 98% for each dNTP

Unit definition : One unit is the amount of enzyme that will incorporate 10 nmol of dNTP into acid-insoluble products in 30 minutes at 74° C with activated salmon sperm DNA as the template-primer.

Reaction mixture for unit definition:

25 mM TAPS (pH 9.3 at 25 °C)

50 mM KCl 2 mM MgCl₂ 0.1 mM DTT

200 μ M each dATP·dGTP·dCTP

100 μ M [³H]-dTTP

0.25 mg/ml activated salmon sperm DNA

Purity : Nicking, endonuclease and exonuclease activity were not detected after the incubation of 0.6 μ g of supercoiled pBR322 DNA, 0.6 μ g of λ DNA or 0.6 μ g of λ -Hind III digest with 10 units of this enzyme for 1 hour at 74°C.

Applications: For DNA amplification by Polymerase Chain Reaction (PCR).

PCR products : As most PCR products amplified with $TaKaRa Ex Taq^{TM}$ have one A added at 3'-termini, the obtained PCR product can be directly used for cloning into T-vector. Also it is possible to clone the product in bluntend vectors after blunting and phosphorylation of the end.

PCR test: Good performance of DNA amplification by PCR was confirmed by using λ DNA as the template (amplified fragment: 20 kb). Good performance of DNA amplification of β -globin gene by PCR was also confirmed by using human genomic DNA as the template (amplified fragment: 17.5 kb).

General reaction mixture for PCR (total 50 μ l)

TaKaRa Ex Taq™ (5 units/ μ I)0.25 μ I10X Ex Taq Buffer5 μ IdNTP Mixture (2.5 mM each)4 μ ITemplate< 500 ng</td>Primer 10.2 - 1.0 μ M (final conc.)Primer 20.2 - 1.0 μ M (final conc.)Sterilized distilled waterup to 50 μ I

PCR condition (an example): When amplifying 1 kb DNA fragment

98℃	10 sec. 30 cycles		98℃ 10		10 sec. –] 20
55℃	30 sec.	30 cycles	or	68℃	1 min	30 cycles
72℃	1 min					

(Note) Denaturation conditions vary depending on the thermal cycler and tubes used for PCR. The recommendation is for 1-10 sec. at 98° C or 10-30 sec. at 94° C.

< Cool Start Method >

"Cool Start Method" + provides more accurate amplification and minimizes amplification of nonspecific bands. This is a simple method that does not require specialized enzymes or additional reagents. Higher reaction specificity can be achieved by combining Hot Start PCR techniques with *Taq* Antibody (Cat. #9002A) and Cool Start method.

Protocol of Cool Start Method

- 1) Keep all reagents on ice until use.
- 2) Prepare the reaction mixture on ice. *1.*2
 - *1: Order of reagent addition does not influence results.
 - *2: Results will not be affected by leaving the mixture on ice for 30 min. before thermal cycling.
- 3) Set a thermal cycler with the designated program. *3
- * 3: PCR conditions dose not need to be changed for Cool Start.
- 4) Set the tubes in a thermal cycler and start thermal cycling immediately.
- +: JAPAN Patent 2576741 for Cool Start Method is owned by SHIMADZU CORPORATION

NOTICE TO PURCHASER: LIMITED LICENSE

[P1] PCR Notice

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[M57] LA Technology

This product is covered by the claims 6-16 of U.S. Patent No. 5,436,149 and its foreign counterpart patent claims.

Note

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