## **Nucleofection Quick Guide**

100 uL reaction 2 ug DNA 1 - 5 million cells/mL

## Do the calculations

- 1. How many cells go into the reaction? You're in luck. I put 500,000 cells in each flask
- 2. How much of your DNA solution?

$$\frac{1 \text{ ng}}{\text{uL}} \times \frac{1000 \text{ uL}}{1 \text{ mL}} \times \frac{1 \text{ ug}}{1000 \text{ng}} = \frac{1 \text{ ug}}{1 \text{ mL}}$$

$$\frac{100 \text{ ug}}{\text{mL}} \times \text{V mL} = 2 \text{ ug}$$

$$\frac{2 \text{ ug}}{100 \text{ ug/mL}} = 0.02 \text{ mL}$$

## Get everything ready

- New flask for nucleofected cells, labeled:
  - o cell type
  - o transfected gene
  - o date
  - o passage #
  - o your initials
- Coverslip-bottom dish, also labeled
- Tube of F10 Hams, labeled with your initials
- Warm PBS, labeled with your initials
- Trypsin (shared)
- Mirus reagent (shared)
- Sterile 2 mL tubes
- Sterile 1.5 mL tubes
- Pipetters and tips
- 5 mL serological pipets
- pipet gun
- cuvette
- dropper
- waste beaker
- spray bottle of ethanol

## Protocol – Keep everything sterile!

- 1. Spray all surfaces. Spray everything that goes into the hood.
- 2. Label new flask to receive nucleofected cells
- 3. 5 mL F10 Hams into flask, 2.5 mL into coverslip dish, move into incubator
- 4. 100 uL Mirus/DNA solution in 1.5 mL tube, into incubator
- 5. Take flask of cells out of incubator, label with your initials
- 6. Use 5 mL serological pipet to remove medium
- 7. Put ~ 1 mL warm PBS into flask (sterile transfer pipet or Pasteur pipet)
- 8. rock to rinse, remove PBS
- 9. Add ~0.5 mL trypsin (KEEP STERILE), rock to coat cells, put in incubator
- 10. Incubate 3 minutes
- 11. Check that cells are in suspension
- 12. Add 1 mL cold F10 Hams (KEEP STERILE) to stop the reaction.
- 13. Mix with transfer pipet
- 14. Transfer ALL to sterile 2 mL tube
- 15. Spin 500 xG 3 minutes
- 16. (BACK TO BSC) Remove supernate
- 17. Resuspend in Mirus/DNA solution
- 18. Put ALL into sterile cuvette, close
- 19. Cuvette to Nucleofector
- 20. Program X001
- 21. Press the button.
- 22. (BACK TO BSC) Use special dropper to transfer one drop to the coverslip dish, the rest to the waiting flask.
- 23. Done!